Inhibitory and Stability Studies on Graphene Oxide Gold Nanozymes

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Submitted: 2024, Sep 16; Accepted: 2024, Oct 07; Published: 2024, Oct 25

Citation: Hormozi Jangi, M. R., Hormozi Jangi, A. R. (2024). Inhibitory and Stability Studies on Graphene Oxide Gold Nanozymes, *Arch of case Rep: Open, 1*(2), 01-06.

Abstract

In this study, peroxidase-like graphene oxide/gold nanoparticles were synthesized and characterized for their biochemical stability including pH and thermal stability by determining their relative activity as an index for their stability monitoring. Besides, the inhibitory effect of metal ions on their enzyme like activity was also studied. The results of thermal stability studies revealed a maximal activity over a wide temperature range of 20-40 °C, revealing high thermal stability of the as-synthesized peroxidase-like graphene oxide/gold nanoparticles. The pH stability measurements revealed a maximal activity over pH= 4.5-5.5 for the as-synthesized peroxidase-like graphene oxide/gold nanoparticles, showing their wide pH range. Moreover, the inhibitory effect of silver ions (as a model metal ion) on their activity was checked at optimal pH and temperature, revealing that the activity of nanozymes was decreased by increasing the concentration of silver ions and reached about 63% upon the addition small portion of silver ions into the reaction media.

Keywords: Inhibitory Effect, Ph Stability, Thermal Stability, Graphene Oxide Gold Nanoparticles

1. Introduction

Despite the high specificity, selectivity, and catalytic performances of the native enzymes, they suffer several disadvantages such as narrow pH and thermal range which limits their practical applications [1-10]. The traditional way for solving these problems is the immobilization of enzymes to enhance their stability and recoverability [10-15]. However, with the fast development of nanoscience in recent years, a wide veracity of nanomaterials with unique spectral, optical, catalytic, and stability has been introduced [16-23]. Among these nanostructures reveal significant enzymelike activity for instance peroxidase-like, oxidase-like, urease-like, and catalase-like, etc. [24-35]. These enzyme-like nanoparticles show significant advantages over native enzymes for instance, high pH and thermal stability, excellent reusability, and high storage stability, making them the best alternative of enzymes for proceeding with enzyme-mediated process [36-44]. Nanozymes have been used for different applications such as dye degradation and the battery industry, as well as, sensing and detection [45-57], especially after the first report of COVID-19 [58, 59], they applied for its clinical sensing [60-67]. Herein, peroxidase-like graphene oxide/gold nanoparticles were synthesized and characterized for their peroxidase-like activity. Considering the significant effect of pH and temperature on the activity of enzyme/nanozymes, the pH and thermal stability of the peroxidase-like graphene oxide/gold

nanoparticles were evaluated by determining their relative activity as an index for their stability monitoring. Besides, the inhibitory effect of silver ions on their activity was checked at optimal pH and temperature.

2. Experimental

2.1. Synthesis of Peroxidase-Like Graphene Oxide/Gold Nanoparticles

For the preparation of graphene oxide gold nanoparticles, the gold nanoparticle and graphene oxide dispersions were mixed with a 1:1 ratio. The reaction mixture was then ultra-sonicated for about 30 minutes to produce the graphene oxide gold nanoparticles. The obtained graphene oxide/gold nanoparticles dispersion was then stored in the dark for further use.

2.2. Enzyme Assay

TMB assay was used for determining the enzyme-like activity of peroxidase-like graphene oxide/gold nanoparticles as a standard method. Briefly, 200 μL of 0.2 mg/mL nanozyme was added to 6.8 mL of 0.2 M acetate buffer (pH 4.5). Then, 1.0 mL TMB (5 mM) and 1.0 mL H2O2 (0.2 M) were added to the solution. After reacting for 10 minutes, the absorbance of the colored product was measured at 650 nm.

3. Results and Discussion

Considering the significant effect of pH and temperature on the activity of enzyme/nanozymes, the pH and thermal stability of the peroxidase-like graphene oxide/gold nanoparticles were evaluated by determining their relative activity as an index for their stability monitoring.

3.1. Effect of pH

The effect of pH on the enzyme-like activity of the as-prepared peroxidase-like graphene oxide/gold nanoparticles was investigated by probing their relative activity by changing the pH of the reaction media. The relative activity of nanozymes was

determined using the following equation; Relative activity = (Activity/Maximal Activity) × 100

The results are shown in Figure 1. This figure shows the relative activity of enzymes as a function of the pH of the media. As can be seen from this figure, the relative activity of nanozymes increased by increasing pH from 3.0, reached the maximal value at 5.0, and then decreased by increasing the pH. It is notable that, the pH stability measurements revealed a maximal activity over pH= 4.5-5.5 for the as-synthesized peroxidase-like graphene oxide gold nanoparticles.

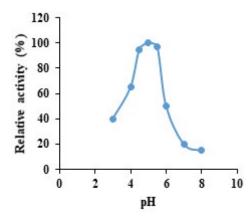


Figure 1: Effect of pH on the enzyme-like activity of the as-prepared peroxidase-like graphene oxide gold nanoparticles

3.2. Effect of Temperature

The effect of temperature on the enzyme-like activity of the asprepared peroxidase-like graphene oxide/gold nanoparticles was investigated by probing their relative activity by changing the operating temperature. The relative activity of nanozymes in different temperatures was determined using the following equation;

Relative activity = $(Activity/Maximal Activity) \times 100$ The results of this investigation are shown in Figure 2. This figure shows the relative activity of the enzyme as a function of the temperature of the reaction media. As can be seen from this figure, the relative activity of nanozymes increased by increasing temperature from 4.0 °C, reached maximal value, and then decreased by increasing the operating temperature. It is notable that, the results of thermal stability studies revealed a maximal activity over a wide temperature range of 20-40 °C, revealing high thermal stability of the as-synthesized peroxidase-like graphene oxide/gold nanoparticles.

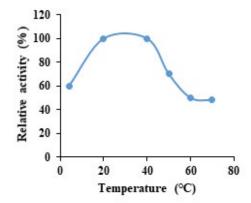


Figure 2: Effect of temperature on the enzyme-like activity of the as-prepared peroxidase-like graphene oxide gold nanoparticles

3.3. Inhibitory Effect of Silver Ions

The inhibitory effect of silver ions on their activity was checked at optimal pH and temperature. To do this, different amounts of silver ions were added into the solution nanozymes, followed by incubation at room temperature for 30.0 min to complete the inhibition process. Thereafter, the enzyme assay was carried out to determine the nanozyme activity. The results were used for

constructing the histogram of nanozyme activity as a function of inhibitor concentration (Figure 3). The results of Figure 3 revealed that the activity of nanozymes was decreased by increasing the concentration of silver ions and reached about 63% upon the addition small portion of silver ions into the reaction media. Hence, silver ions can significantly inhibit the enzyme-like activity of the as-prepared peroxidase-like graphene oxide/gold nanoparticles.

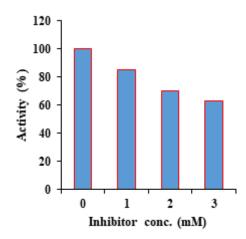


Figure 3: Inhibitory effect of silver ions on the enzyme-like activity of the as-prepared peroxidase-like graphene oxide gold nanoparticles.

4. Conclusions

Herein, peroxidase-like graphene oxide/gold nanoparticles were synthesized and characterized for their peroxidase-like activity. Considering the significant effect of pH and temperature on the activity of enzyme/nanozymes, the pH and thermal stability of the peroxidase-like graphene oxide/gold nanoparticles were evaluated by determining their relative activity as an index for their stability monitoring. The results of thermal stability studies revealed a maximal activity over a wide temperature range of 20-40 °C, revealing high thermal stability of the as-synthesized peroxidaselike graphene oxide/gold nanoparticles. Besides, the pH stability measurements revealed a maximal activity over pH= 4.5-5.5 for the as-synthesized peroxidase-like graphene oxide/gold nanoparticles. Moreover, the inhibitory effect of silver ions on their activity was checked at optimal pH and temperature, revealing that the activity of nanozymes was decreased by increasing the concentration of silver ions and reached about 63% upon the addition small portion of silver ions into the reaction media.

Acknowledgment

None.

Conflict of interest

None.

References

- 1. Uhlig, H. (Ed.). (1998). Industrial enzymes and their applications. John Wiley & Sons.
- 2. Sizer, I. W. (1964). Enzymes and their applications. In Advances in Applied Microbiology (Vol. 6, pp. 207-226).

Academic Press.

- 3. Underkofler, L. A., Barton, R. R., & Rennert, S. S. (1958). Production of microbial enzymes and their applications. *Applied microbiology*, *6*(3), 212-221.
- 4. Jangi, S. R. H. A. (2023). Mini-Review on Nanozyme Chemistry with Focus on Analytical and Bioanalytical Sensing Applications.
- Jangi, S. R. H. (2023). Determining kinetics parameters of bovine serum albumin-protected gold nanozymes toward different substrates. *Qeios*.
- Ghaffari-Moghaddam, M., Eslahi, H., Omay, D., & Zakipour-Rahimabadi, E. (2014). Industrial applications of enzymes. Review Journal of Chemistry, 4, 341-361.
- 7. Hormozi Jangi, S. R. (2023). Naked-Eye Sensing of SARS-CoV-2 Utilizing Nanozymatic Nanoassays. *J Pediatr Neonatal Biol*, 8(4), 283-289.
- 8. Hormozi Jangi, S. R. (2023). BSA-Stabilized Gold-Nanozymes Reveal 4-Order Higher Catalytic Efficiency and 2-Fold Higher Substrate Affinity than Mno2-Nanozymes. *J App Mat Sci & Engg Res*, 7(2), 166-171.
- 9. Jangi, S. R. H. (2023). Time Course Evaluation of Nanozyme-Mediated Reversible/Irreversible Oxidation Reactions over Silver Nanoparticles as Peroxidase Alternatives.
- 10. Hormozi Jangi, S. R. (2023). A Brief Overview of Nanozyme-Based Colorimetric and Fluorometric Sensors for Early Diagnosis of COVID-19. *Trans Med OA*, 1(2), 76-84.
- 11. Jangi, S. R. H., Akhond, M., & Dehghani, Z. (2020). High throughput covalent immobilization process for improvement of shelf-life, operational cycles, relative activity in organic media and enzymatic kinetics of urease and its application for

- urea removal from water samples. *Process Biochemistry*, 90, 102-112
- 12. Jangi, S. R. H., & Akhond, M. (2021). High throughput urease immobilization onto a new metal-organic framework called nanosized electroactive quasi-coral-340 (NEQC-340) for water treatment and safe blood cleaning. *Process Biochemistry*, 105, 79-90.
- 13. Liu, D. M., & Dong, C. (2020). Recent advances in nanocarrier immobilized enzymes and their applications. *Process Biochemistry*, *92*, 464-475.
- 14. Jangi, S. R. H., & Akhond, M. (2022). Introducing a covalent thiol-based protected immobilized acetylcholinesterase with enhanced enzymatic performances for biosynthesis of esters. *Process Biochemistry*, 120, 138-155.
- Cacicedo, M. L., Manzo, R. M., Municoy, S., Bonazza, H. L., & Islan, G. A., et al. (2019). Immobilized enzymes and their applications. In *Advances in enzyme technology* (pp. 169-200). Elsevier.
- 16. Jangi, S. R. H. (2024). Developing a label-free full-range highly selective pH nanobiosensor using a novel high quantum yield pH-responsive activated-protein-protected gold nanocluster prepared by a novel ultrasonication-proteinassisted procedure. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, 322, 124829.
- Jangi, S. R. H. (2024). Developing a novel ultraselective and ultrasensitive label-free direct spectrofluorimetric nanobiosensor for direct highly fast field detection of explosive triacetone triperoxide. *Analytica Chimica Acta*, 1320, 343016.
- 18. Dehghani, Z., Akhond, M., Jangi, S. R. H., & Absalan, G. (2024). Highly sensitive enantioselective spectrofluorimetric determination of R-/S-mandelic acid using l-tryptophan-modified amino-functional silica-coated N-doped carbon dots as novel high-throughput chiral nanoprobes. *Talanta*, 266, 124977.
- Hormozi Jangi, S. R., & Gholamhosseinzadeh, E. (2023). Developing an ultra-reproducible and ultrasensitive label-free nanoassay for L-methionine quantification in biological samples toward application in homocystinuria diagnosis. Chemical Papers, 77(11), 6505-6517.
- 20. Hormozi Jangi, S. R. (2023). Low-temperature destructive hydrodechlorination of long-chain chlorinated paraffins to diesel and gasoline range hydrocarbons over a novel low-cost reusable ZSM-5@ Al-MCM nanocatalyst: a new approach toward reuse instead of common mineralization. *Chemical Papers*, 77(9), 4963-4977.
- Hormozi Jangi, S. R., & Akhond, M. (2023). Evaluating effect of solvent type, metal-core salt and heat-treatment on organic content, yield, and coordination strength in magnesium-based metal-organic frameworks using elemental analysis and Fourier transform infrared spectroscopy. *Iranian Journal of Chemistry*, 5(2), 225-234.
- 22. Jangi, S. R. H., & Akhond, M. (2021). Ultrasensitive label-free enantioselective quantification of d-/l-leucine enantiomers with a novel detection mechanism using an ultra-small high-quantum yield N-doped CDs prepared by a novel highly fast

- solvent-free method. Sensors and Actuators B: Chemical, 339, 129901.
- 23. HORMOZI JANGI, S. R., & Akhond, M. (2020). High throughput green reduction of tris (p-nitrophenyl) amine at ambient temperature over homogenous AgNPs as H-transfer catalyst. *Journal of Chemical Sciences*, *132*, 1-8.
- Wang, G. L., Zhu, X. Y., Jiao, H. J., Dong, Y. M., & Li, Z. J. (2012). Ultrasensitive and dual functional colorimetric sensors for mercury (II) ions and hydrogen peroxide based on catalytic reduction property of silver nanoparticles. *Biosensors and Bioelectronics*, 31(1), 337-342.
- 25. Tan, L., Zhang, Y., Qiang, H., Li, Y., Sun, J., Hu, L., & Chen, Z. (2016). A sensitive Hg (II) colorimetric sensor based on synergistic catalytic effect of gold nanoparticles and Hg. *Sensors and Actuators B: Chemical*, 229, 686-691.
- 26. Akhond, M., Hormozi Jangi, S. R., Barzegar, S., & Absalan, G. (2020). Introducing a nanozyme-based sensor for selective and sensitive detection of mercury (II) using its inhibiting effect on production of an indamine polymer through a stable n-electron irreversible system. *Chemical Papers*, 74, 1321-1330.
- 27. Jangi, S. R. H., Akhond, M., & Absalan, G. (2020). A novel selective and sensitive multinanozyme colorimetric method for glutathione detection by using an indamine polymer. *Analytica Chimica Acta, 1127*, 1-8.
- 28. Hormozi Jangi, S. R., Akhond, M., & Absalan, G. (2020). A field-applicable colorimetric assay for notorious explosive triacetone triperoxide through nanozyme-catalyzed irreversible oxidation of 3, 3'-diaminobenzidine. *Microchimica Acta, 187*, 1-10.
- 29. Jangi, S. R. H., & Akhond, M. (2020). Synthesis and characterization of a novel metal-organic framework called nanosized electroactive quasi-coral-340 (NEQC-340) and its application for constructing a reusable nanozyme-based sensor for selective and sensitive glutathione quantification. *Microchemical Journal*, 158, 105328.
- Wang, J., Li, W., & Zheng, Y. Q. (2019). Nitro-functionalized metal—organic frameworks with catalase mimic properties for glutathione detection. *Analyst*, 144(20), 6041-6047.
- 31. Khoshsafar, H., Karimian, N., Nguyen, T. A., Fakhri, H., Khanmohammadi, A., Hajian, A., & Bagheri, H. (2022). Enzymeless voltammetric sensor for simultaneous determination of parathion and paraoxon based on Nd-based metal-organic framework. *Chemosphere*, 292, 133440.
- 32. Kaimal, R., Dube, A., Al Souwaileh, A., Wu, J. J., & Anandan, S. (2024). A copper metal-organic framework-based electrochemical sensor for identification of glutathione in pharmaceutical samples. *Analyst*, 149(3), 947-957.
- 33. Diao, Q., Chen, X., Tang, Z., Li, S., Tian, Q., Bu, Z., ... & Niu, X. (2024). Nanozymes: powerful catalytic materials for environmental pollutant detection and degradation. *Environmental Science: Nano*.
- 34. Hormozi Jangi, S. R., & Dehghani, Z. (2023). Kinetics and biochemical characterization of silver nanozymes and investigating impact of storage conditions on their activity

- and shelf-life. Chemical Research and Nanomaterials, 1(4), 25-33.
- 35. Hormozi Jangi, S. R. (2023). Synthesis and characterization of magnesium-based metal-organic frameworks and investigating the effect of coordination solvent on their biocompatibility. *Chemical Research and Nanomaterials*, *1*(4), 1-9.
- 36. Hormozi Jangi, S. R. (2023). Evaluation of Biochemical Behavior and Stability of Gold Nanoparticles with High Intrinsic Peroxidase-Like Activity. *Petro Chem Indus Intern*, 6(4), 234-239.
- 37. Jangi, S. R. H. (2023). Biochemical characterization of enzyme-like silver nanoparticles toward nanozyme-catalysed oxidation reactions. *Micromaterials and Interfaces*, *1*(1).
- 38. Jangi, S. R. H. (2023). Effect of daylight and air oxygen on nanozymatic activity of unmodified silver nanoparticles: Shelf-stability. *Qeios*.
- 39. Lou-Franco, J., Das, B., Elliott, C., & Cao, C. (2021). Gold nanozymes: from concept to biomedical applications. *Nano-Micro Letters*, *13*, 1-36.
- 40. Hormozi Jangi, S. R. (2023). A Comparative Study on Kinetics Performances of BSA-gold Nanozymes for Nanozymemediat-ed Oxidation of 3, 3', 5, 5'-Tetramethylbenzidine and 3, 3'-Diaminobenzidine. *Biochem Mol Biol J, 9*, 21.
- 41. Abdel-Lateef, M. A. (2022). Utilization of the peroxidase-like activity of silver nanoparticles nanozyme on O-phenylenediamine/H2O2 system for fluorescence detection of mercury (II) ions. *Scientific Reports*, 12(1), 6953.
- 42. Hormozi Jangi, S. R. (2023). Experimental Evaluation of Kinetic Characteristics of SiO2@ AuNPs Nanocom-posite and BSA-stabilized gold Nanoparticles toward Peroxidase-Mediated Reactions. *Adv Nanoscie Nanotec*, 7(1), 01-11.
- 43. Jangi, S. R. H. (2023). Experimental evaluation of kinetics and biochemical characteristics of MnO2 nanoparticles as high throughput peroxidase-mimetic nanomaterials. *Micromaterials and Interfaces*, *1*(1).
- 44. Hormozi Jangi, S. R., & Dehghani, Z. (2024). Captopril Molecules Reveal Strong pH-, Temperature-, And Concentration-Dependent Inhibitory Effect on Nanozymatic Activity of Peroxidase-Like Nitrogen-Doped Carbon Dots. *Petro Chem Indus Intern*, 7(1), 01-07.
- 45. Hormozi Jangi, S. R., & Dehghani, Z. (2023). Spectrophotometric quantification of hydrogen peroxide utilizing silver nanozyme. *Chemical Research and Nanomaterials*, 2(1), 15-23.
- 46. Jangi, A. R. H., Jangi, M. R. H., & Jangi, S. R. H. (2020). Detection mechanism and classification of design principles of peroxidase mimic based colorimetric sensors: A brief overview. *Chinese Journal of Chemical Engineering*, 28(6), 1492-1503.
- 47. Hormozi Jangi, S. R. (2023). Detection mechanism and principles of the multinanozyme systems as the new generation of nanozyme-mediated sensing assays: A critical review. *Petro Chem Indus Intern*, 6(5), 349-357.
- 48. Ahmadi-Leilakouhi, B., Hormozi Jangi, S. R., & Khorshidi,

- A. (2023). Introducing a novel photo-induced nanozymatic method for high throughput reusable biodegradation of organic dyes. *Chemical Papers*, 77(2), 1033-1046.
- 49. Jangi, S. R. H., Davoudli, H. K., Delshad, Y., Jangi, M. R. H., & Jangi, A. R. H. (2020). A novel and reusable multinanozyme system for sensitive and selective quantification of hydrogen peroxide and highly efficient degradation of organic dye. *Surfaces and Interfaces*, 21, 100771.
- 50. Hormozi Jangi, S. R. (2024). Lithium-electroactive peroxidaselike MnO2 nanomaterials as an ultrasensitive and selective sensing platform for carcinogenic 3, 3'-diaminobenzidine and high-capacity lithium-ion batteries cathode materials. *Chemical Papers*, 1-13.
- 51. Jangi, S. R. H. (2023). Introducing a high throughput nanozymatic method for eco-friendly nanozyme-mediated degradation of methylene blue in real water media. *Sustainable Chemical Engineering*, 90-99.
- 52. Huang, Y., Ren, J., & Qu, X. (2019). Nanozymes: classification, catalytic mechanisms, activity regulation, and applications. *Chemical reviews*, 119(6), 4357-4412.
- 53. Liang, M., & Yan, X. (2019). Nanozymes: from new concepts, mechanisms, and standards to applications. *Accounts of chemical research*, 52(8), 2190-2200.
- 54. Ai, Y., Hu, Z. N., Liang, X., Sun, H. B., & Xin, H., et al. (2022). Recent advances in nanozymes: from matters to bioapplications. *Advanced Functional Materials*, 32(14), 2110432.
- 55. Wu, J., Wang, X., Wang, Q., Lou, Z., & Li, S., et al. (2019). Nanomaterials with enzyme-like characteristics (nanozymes): next-generation artificial enzymes (II). *Chemical Society Reviews*, 48(4), 1004-1076.
- 56. Meng, Y., Li, W., Pan, X., & Gadd, G. M. (2020). Applications of nanozymes in the environment. *Environmental Science: Nano*, 7(5), 1305-1318.
- 57. Dong, H., Fan, Y., Zhang, W., Gu, N., & Zhang, Y. (2019). Catalytic mechanisms of nanozymes and their applications in biomedicine. *Bioconjugate chemistry*, 30(5), 1273-1296.
- 58. Jangi S. R., H. (2023). Natural Polyphenols of Pomegranate and Black Tea Juices can Combat COVID-19 through their SARS-CoV-2 3C-like Protease-inhibitory Activity.
- 59. Hormozi Jangi, S. R. (2023). A brief overview on clinical and epidemiological features, mechanism of action, and diagnosis of novel global pandemic infectious disease, Covid-19, and its comparison with Sars, Mers, And H1n1. *World J Clin Med Img*, 2(1), 45-52.
- 60. Ali, J., Elahi, S. N., Ali, A., Waseem, H., & Abid, R., et al. (2021). Unveiling the potential role of nanozymes in combating the COVID-19 outbreak. *Nanomaterials*, 11(5), 1328.
- 61. Li, D., Zhao, B., Zhuang, P., & Mei, X. (2023). Development of nanozymes for promising alleviation of COVID-19-associated arthritis. *Biomaterials Science*.
- 62. Kumawat, M., Umapathi, A., Lichtfouse, E., & Daima, H. K. (2021). Nanozymes to fight the COVID-19 and future pandemics. *Environmental Chemistry Letters*, 19(6), 3951-

- 3957.
- 63. Wang, J., Xie, Q., Song, H., Chen, X., & Zhang, X., et al. (2023). Utilizing nanozymes for combating COVID-19: advancements in diagnostics, treatments, and preventative measures. *Journal of Nanobiotechnology*, 21(1), 200.
- 64. Ali, G. K., & Omer, K. M. (2022). Nanozyme and stimulated fluorescent Cu-based metal—organic frameworks (Cu-MOFs) functionalized with engineered aptamers as a molecular recognition element for thrombin detection in the plasma of COVID-19 patients. *ACS omega*, 7(41), 36804-36810.
- 65. Lee, J., Liao, H., Wang, Q., Han, J., & Han, J. H., et al. (2022,

- February). Exploration of nanozymes in viral diagnosis and therapy. In *Exploration* (Vol. 2, No. 1, p. 20210086).
- Liu, D., Ju, C., Han, C., Shi, R., Chen, X., Duan, D., ... & Yan, X. (2021). Nanozyme chemiluminescence paper test for rapid and sensitive detection of SARS-CoV-2 antigen. *Biosensors and Bioelectronics*, 173, 112817.
- 67. Wang, D., Zhang, B., Ding, H., Liu, D., & Xiang, J., et al. (2021). TiO2 supported single Ag atoms nanozyme for elimination of SARS-CoV2. *Nano Today*, 40, 101243.

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