

Research Article

Translational Medicine : Open Access

Advancing Precision Drug Therapy in Pregnant Women: PBPK Modeling of Antiviral Drugs

Mohamed Abdulsamed*, Ashraf A Naass, Mohamed S A Eswani, Mohamed O Elbasir, and Sedigh Bashir

Libyan Biotechnology Research Center

*Corresponding Author Mohamed Abdulsamed, Libyan Biotechnology Research Center.

Submitted: 2024, Aug 23; Accepted: 2024, Sep 12; Published: 2024, Sep 25

Citation: Abdulsamed, M., Naass, A. A., Eswani, M. S. A., Elbasir, M. O., Bashir. S. (2024). Advancing Precision Drug Therapy in Pregnant Women: PBPK Modeling of Antiviral Drugs. *Trans Med OA*, 2(2), 01-17.

Abstract

PBPK/PD modeling is essential in modern drug development. Traditional drug development methods frequently rely on trial and error, which can be time-consuming, costly, and could be risky. Predicting pharmacokinetics (PK) of drugs in pregnant women, encompassing the intricate aspect of placental drug transfer, remains a complex task. This study was to compare of simulated or predicted and observed (previously published approaches) pharmacokinetic parameters among the four antiviral drugs in pregnant and non-pregnant women. In addition, this investigation endeavors to construct and assess physiologically-based pharmacokinetic (PBPK) models specific to maternal-fetal interactions for four antiviral drugs, Acyclovir, Emtricitabine, Dolutegravir (DTG) and Raltegravir (RAL). PBPK models were built with the Open Systems Pharmacology software suite (PK-Sim/MoBi). Different approaches to inform placental drug transfer were applied and compared. Model performance was evaluated using in vivo all 4 a forementioned antiviral maternal plasma concentrations during the 2nd and 3rd trimesters and umbilical vein concentrations at delivery. All clinical in vivo data were obtained from the International Maternal paediatric and Adolescent AIDS Clinical Trials (IMPAACT) Network P1026s study. The PBPK models successfully predicted plasma concentration-time profiles of four antiviral drugs in the 2nd and 3rd trimesters and most predicted PK parameters fell within a 1.33-fold error range. Predicted umbilical vein concentrations of DTG among others were in reasonable agreement with in vivo data but were sensitive to changes in the placental partition coefficient and transplacental clearance. Maternal-fetal PBPK modeling reliably predicted maternal PK of previously mentioned antiviral during pregnancy. For the fetal PK, data on the unbound fraction of highly protein-bound DTG has proven to be important to adequately capture changes in total clearance in silico. More research efforts, along with clinical data, are needed to verify the predictions of fetal PK of antiviral. In conclusion, the findings suggest the feasibility of employing physiologically-based pharmacokinetic (PBPK) models to assess the disposition of antiviral drugs in pregnant women and their fetuses.

Keywords: Pharmacokinetic, Modeling, Pregnancy, Antiviral Drugs, PBPK. Maternal-Fetal

1. Introduction

Medication utilization during pregnancy is widespread and increasing. In a prospective, longitudinal cohort study focusing on prescription drugs and other medication consumption during pregnancy, findings indicate that 97.1% of women included in the study engaged in the use of at least one medication throughout their pregnancy, and 30.5% women took 5 or more medications [1]. Although physiological changes during pregnancy can notably influence drug disposition, the inclusion of pregnant women in clinical studies remains limited. Consequently, clinicians frequently encounter the challenge of prescribing medications during pregnancy without access to specific information regarding pharmacokinetics (PK) and safety in this population. Particularly, antiviral medications are commonly administered during pregnancy for both maternal treatment and prophylaxis to mitigate the risk of perinatal viral transmission. Acyclovir, an antiviral drug effective against the herpes simplex

virus, stands as a noteworthy example. Herpes simplex virus constitutes one of the most prevalent sexually transmitted infections, bearing the potential for neonatal morbidity or mortality if infection during the neonatal period is not averted or promptly addressed [2]. Emtricitabine, Dolutegravir, and Raltegravir represent antiretroviral drugs with efficacy against human immunodeficiency virus (HIV) infection. As of 2016, a substantial cohort of 19.5 million individuals living with HIV is actively receiving antiviral treatment [3]. With the expanding depth of knowledge concerning anatomical and physiological alterations during pregnancy, the application of physiologically based pharmacokinetic (PBPK) models to extant pharmacokinetic (PK) data for pregnant women has become feasible, enhancing confidence in these models. Leveraging their mechanistic foundation, PBPK models offer valuable insights into the physiological mechanisms underpinning PK changes. Anticipating PK alterations in specific populations,

such as pregnant women, prior to the commencement of clinical trials, can streamline the design and execution of such studies. However, a prerequisite for such applications is a robust confidence level in the established PBPK model. While several PBPK models for pregnancy have been established [4]. a notable gap persists, as many predominantly focus on maternal PK changes, with limited consideration for drug exposure in the fetal compartment [5]. According to the collected data This study presents the development of maternal-fetal PBPK models for group of antiviral drugs which are acyclovir, Emtricitabine, Dolutegravir and Raltegravir. The objectives of this study were to compare of simulated or predicted and observed (previously published approaches) pharmacokinetic parameters among the four antiviral drugs in pregnant and non-pregnant women and to evaluate the model predictions of the PK in the mother at different stages of pregnancy.

2. Methodology

The study was based on data collection through PubMed, Scopus and Google scholar using the medical terms "Physiologically based pharmacokinetic pregnancy modeling" and Antiretroviral drugs. These obtained data and literature were focused on using open systems pharmacology software package version 8.0 (https://www.open-systems-pharmacology.org/).(PK-Sim) and MoBi. In addition, R Foundation for stastical computing version 3.4.1 software. On other hand, all clinical in vivo data were obtained from the international maternal pediatric and adolescent AIDS clinical trial (IMPAACT) Network P 1026s study. All sources code and the model developed available on GitHub accessible via(https://www.open-systems-pharmacology.org/).

3. General Workflow

The procedural framework governing the advancement of the pregnancy Physiologically-Based Pharmacokinetic (PBPK) model has been exhaustively explicated in a previous scholarly account [6]. The schematic depiction of this methodology is presented in Figure 1. Briefly, a PBPK model was initially developed for a virtual non-pregnant population and evaluated by comparing simulation results with the observed in vivo PK data in non-pregnant subjects reported in the comparison studies. Thereafter, the non-pregnant PBPK model was translated to pregnancy by substituting the standard model structure with the pregnancy structure and parametrizing the model for the respective gestational age as described before[6]. PK predictions in pregnant women were evaluated by comparison with in vivo PK data obtained from clinical trials of IMPAACT P1026s.

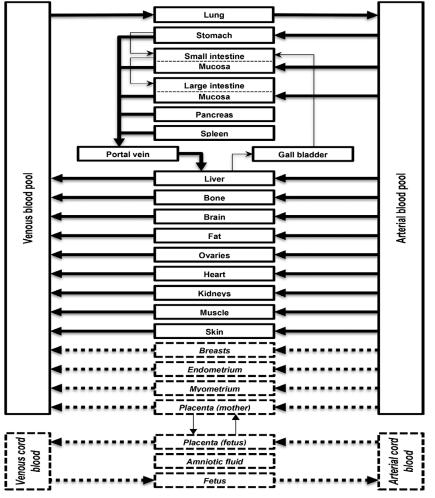


Figure 1: Pregnancy PBPK Model Structure. Thick Arrows Represent Drug Transport Via Blood Flow, and Thin Arrows Via Other Pathways (eg, Via Passage in the Gastrointestinal Tract, Biliary Excretion through the Gallbladder, and Diffusive Transfer in the Placenta). Compartments that are Exclusively Part of the Pregnancy PBPK Model Structure are Shown in Italics with Dashed Borders and Dashed Arrows for Drug Transfer Via the Blood Flow [7].

4. Development of PBPK Models 4.1 Acyclovir

Demonstrates renal excretion efficiency after intravenous administration, with a range of 61% to 91% of the radioactively labeled dose being excreted unchanged in urine (corrected for the amount of radioactivity lost), and 8.5% to 14.1% is metabolized to CMMG (9-carboxy methoxymethylguanine) [8]. OAT (organic anion transporter) 2 has been suggested to be the main transporter involved in renal secretion [9]. CMMG is formed in a 2-step reaction involving a reversible oxidation catalyzed by alcohol dehydrogenase and subsequent irreversible transformation to CMMG via aldehyde dehydrogenase 2 [10,11]. Pharmacokinetic profiles in nonpregnant subjects were obtained from a study by Laskin et al, who investigated acyclovir disposition after intravenous administration of different doses, and additionally from another study that investigated acyclovir PK after oral administration of 400 mg acyclovir as either a suspension or a tablet [12,13]. Two studies, encompassing pregnant women nearing full term, have presented maternal plasma concentrationtime data subsequent to the administration of 400 mg acvclovir. both in single and multiple dosage scenarios. and were used for evaluating the predicted maternal plasma concentrations. Another study reported paired concentration measurements in the maternal plasma and in the umbilical vein obtained at delivery, which were used for evaluating the predicted concentrations in the umbilical vein blood compartment of the model [14-16].

4.2 Emtricitabine

Is a nucleoside reverse transcriptase inhibitor with a daily oral dose of 200 mg in both pregnant and non-pregnant adult populations [17]. Emtricitabine undergoes predominant elimination in an unchanged form through renal excretion, involving a combination of glomerular filtration and tubular secretion, accounting for 71% of the radioactive dose (adjusted for lost radioactivity). PK simulations in the non-pregnant population were evaluated by comparison with in vivo data obtained from 5 different studies that investigated the PK of Emtricitabine in non-pregnant subjects after single and multiple oral administration of 200 mg [18-20]. In pregnant women the PK was predicted in different populations, namely in 3 different gestational age groups of non-laboring pregnant women (23-30, 31-35, and 36-42 gestational weeks) and in women in labor between 34 and 39 weeks of gestation. Drug concentrations in the umbilical vein were predicted in the latter group. PK predictions in pregnant populations were evaluated through comparison with some hitherto unpublished and published in vivo data [21]. The clinical in vivo data were from the IMPAACT (International Maternal Pediatric and Adolescent AIDS Clinical Trials) Network study P1026s. In this clinical study steady-state PK samples were collected at 0, 1, 2, 4, 6, 8, 12, and 24 hours after dosing. The protocol for this study was approved by the responsible institutional review boards [22].

4.3 Dolutegravir

Is prescribed at a daily dosage of 50 mg in the form of an orally administered tablet, administered once daily to both pregnant and non-pregnant adult patients who are treatment-naïve or treatment-experienced, and lack resistance to integrase strand transfer inhibitors (INSTIs). The predominant elimination pathway for Dolutegravir involves metabolic processes facilitated by various enzymes, namely UGT1A1, UGT1A3, UGT1A9, and CYP3A4, accounting for approximately 51%, 2.8%, 5.5%, and 21% of the administered dose, respectively [23]. In the developed model, the contribution of UGT1A3 and 1A9 to total glucuronidation was combined into the biotransformation pathway mediated by UGT1A1. Finally, to obtain dose fractions summing up to 1.0, the dose fraction metabolized via UGT1A1 was increased to 0.79 based on the assumption that the reported value (0.51) may be underestimated due to hydrolyzation and back conversion of the glucuronide to DTG in the feces, as discussed elsewhere [23]. In the pregnancy PBPK model, physiologic parameters were adjusted to the respective stage of pregnancy [24]. Additionally, the reference concentrations of UGT1A1 and CYP3A4 (quantifying the concentrations of these enzymes in the model) were increased to reflect induction of these enzymes. Specifically, CYP3A4 reference concentration was increased by a factor of 1.60 in the 2nd and 3rd trimesters and UGT1A1 reference concentration by a factor of 1.75 in the 2nd trimester and 1.92 in the 3rd trimester [25,26]. The fraction unbound of DTG, averaging 0.0070 in non-pregnant subjects, was also adjusted based on the albumin concentration measured in the herein investigated study subjects and I [27]. Specifically, the mean albumin concentration measured in the 2nd trimester, 3rd trimester and 6 – 12 weeks postpartum was 34.4 g/L, 32.8 g/L and 41.4 g/L, respectively. Using a previously presented scaling approach [24]. these measurements resulted in a fraction unbound of 0.0084 and 0.0088 in the 2nd and 3rd trimester, respectively. PK simulations in the non-pregnant population were evaluated by comparison with in vivo data obtained from eight clinical studies reported in the literature that investigated the PK of DTG in a total of 22 different groups of non-pregnant subjects after single and multiple oral administrations of 2 to 100 mg as granule suspension or 50 mg as tablet in fasted or fed state. In pregnant women, the PK was predicted in 2 different gestational age groups of non-laboring pregnant women in the 2nd trimester (median gestational age [range]: 23.5 weeks) and 3rd trimester (median gestational age [range]: 33 weeks), and in women in labor (median gestational age [range]: 38 weeks) [31-36, 38-50]. Drug concentrations in the blood plasma of the umbilical vein were predicted in the laboring pregnant women group.

4.4 Raltegravir

Is prescribed in either 400 mg twice daily or 1200 mg once daily oral tablet regimens for adult patients, encompassing both pregnant and non-pregnant individuals who are treatment-naive or treatment-experienced. The predominant elimination pathway for raltegravir involves metabolism catalyzed by UGT1A1 and UGT1A9, constituting approximately 70% and 11% of the administered dose, respectively.37. Additionally, approximately 9% is eliminated unchanged through the kidneys [37]. The non-pregnant PBPK model for RAL was obtained from the OSP GitHub repository (https://github.com/Open-Systems-Pharmacology/Raltegravir-Model/releases) where an extensive description and evaluation of the model can be found. In the pregnancy PBPK model, physiologic parameters and the reference concentrations of UGT1A1 were adjusted to the respective stage of pregnancy as described above. Since no information on the effect of pregnancy on UGT1A9 could be found, this enzyme was not induced in the presented model. Similar to DTG, the fraction unbound of RAL, averaging 0.17 in non-pregnant adultsm,38 was adjusted based on the mean albumin concentration measured in the 2nd trimester, 3rd trimester and 6 - 12weeks postpartum (34.1 g/L, 32.4 g/L and 41.4 g/L, respectively) resulting in a fraction unbound of 0.198 and 0.206 in the 2nd and 3 rd trimester, respectively [51]. Additional information on model development and translation to pregnancy can be found in the Supplemental Material. In pregnant women, the PK were predicted in 2 different gestational age groups of non-laboring pregnant women in the 2nd trimester (median gestational age [range]: 23.5 31-36 weeks) and the 3rd trimester (median gestational age [range]: 34 41-49 weeks), and in women in labor (median gestational age [range]: 38 47-51 weeks). Drug concentrations in the blood plasma of the umbilical vein were predicted in the laboring pregnant women group.

4.5 Evaluation of PBPK Models

The PBPK models were evaluated through visual comparison of observed in vivo plasma concentration-time profiles with the concentrations simulated in non-pregnant women or predicted in pregnant women. Other visual predictive checks included goodness-of-fit (GOF) plots, in which individual in vivo concentration values, if available, were combined at each time point to geometric mean values. Additionally, simulated or predicted PK parameters were compared with observed PK parameters obtained from the mean in vivo plasma concentration time profiles. Ratios of simulated or predicted to observed PK parameters were also estimated.

5. Results

5.1 Non-pregnant and Pregnant PBPK Models 5.1.1 Acyclovir and Emtricitabine

The outcomes detailed herein exclusively apply to female who are not currently pregnant. The Figure 2 illustrates the Simulated of plasma concentration-time profiles of acyclovir subsequent to intravenous administration. In this Figure, the paragraphs presented below depict the Goodness-of-fit plot for plasma concentrations of acyclovir (upper panels) and Emtricitabine (lower panels) in non-pregnant subjects (left panels) and pregnant women (right panels). The continuous line denotes the line of identity, while the dashed lines delineate the 2-fold error range. In the upper left panel, acyclovir plasma concentrations are presented for non-pregnant subjects, with black circles representing concentrations for the suspension and gray circles signifying concentrations in steady state for the suspension. Meanwhile, in the upper right panel, acyclovir plasma concentrations in pregnant women are illustrated, with black circles indicating concentrations after a single dose and gray circles representing concentrations in steady state. Lower left panel: Emtricitabine plasma concentrations in non-pregnant subjects; black circles indicate the concentrations after a single dose, gray circles indicate concentrations at steady state, gray squares indicate concentrations after a single dose, black squares indicate concentrations after a single dose, and black triangles indicate concentrations at steady state. Lower right panel: Plasma concentrations of Emtricitabine in pregnant women are delineated as follows: black circles denote concentrations in women at gestational age 23-30 weeks, gray circles represent concentrations in women at gestational age 31-35 weeks, and black squares indicate concentrations in women at gestational age 36-42 weeks [13-21].

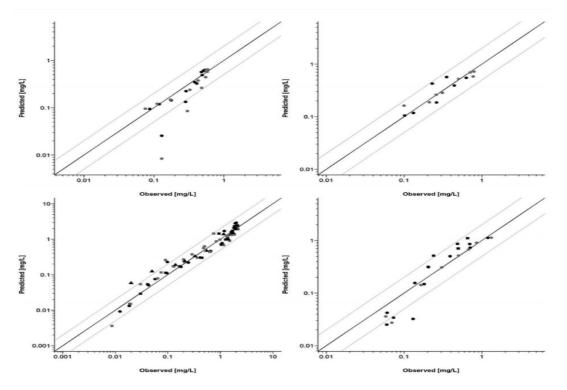


Figure 2: Goodness-of-Fit Plot for Plasma Concentrations of Acyclovir (upper panels) and Emtricitabine (lower panels) in Non-Pregnant Subjects (left panels) and Pregnant Women (right panels).

5.1.2 Evaluation of Predicted Drug Pharmacokinetics for Pregnant Women

The results described in the following refer exclusively to pregnant women. Figure 3 (panels A and B) and Figure 4 (panels A, B, and C) show the anticipated maternal plasma concentration-time profiles of acyclovir and emtricitabine were consistent with the observed in vivo data, demonstrating good agreement between predictions and actual outcomes. Figure 2 presents the predicted mean concentration values in a GOF plot and indicates that all maternal concentrations of acyclovir were predicted within a 2-fold error range, whereas for emtricitabine 79% of the concentration values were predicted within that range. PK parameters calculated from the predicted emtricitabine and acyclovir plasma concentration-time profiles are compared with the observed in vivo PK parameters in Table 2. For acyclovir, the ratios of predicted to observed PK parameters were all within a 25% error range, whereas for emtricitabine most of these ratios were within that range.

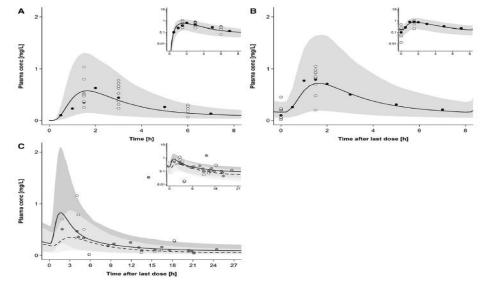


Figure 3: Plasma concentration-time profiles of acyclovir following oral administration of 400 mg in pregnant women. Semi-log scale figures are given as inset figures in the top right corners. Observed in vivo data were taken from published studies. **A graph shows,** Single dose in pregnant women with an average gestational age of 36 weeks.

B graph shows, Multiple doses in steady state in pregnant women with an average gestational age of 38 weeks.

C graph shows, Multiple doses in steady state in pregnant women with an average gestational age of 40 weeks [14-16].

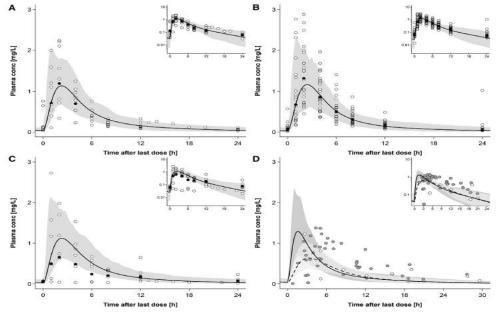


Figure 4: Plasma Concentration-Time Profiles of Emtricitabine following Oral Administration of 200 mg in Pregnant Women in Steady State. Semi-Log Scale Figures are Given as Inset Figures in the Top Right Corners.

A, Pregnant women with a gestational age of 23-30 weeks.

- B, Pregnant women with a gestational age of 31-35 weeks.
- C, Pregnant women with a gestational age of 36-42 weeks.

D, represents individual concentration data in maternal plasma and umbilical vein [2,21].

Trans Med OA, 2024

6. Non-pregnant and Pregnant PBPK Models 6.1 Dolutegravir and Raltegravir

6.1.1 Dolutegravir

The simulated of plasma concentration-time profiles of DTG in non-pregnant populations result from simulation following the administration of a 50 mg tablet once daily in a fed state,

mirroring the dosing regimen employed in pregnant women (Figure 5). Additionally, the figure shown simulated plasma concentration-time profiles for various dosing regimens. Table 1 provides the ratios of simulated to observed pharmacokinetic (PK) parameters in non-pregnant subjects, presenting both the absolute simulated and observed values for these parameters.

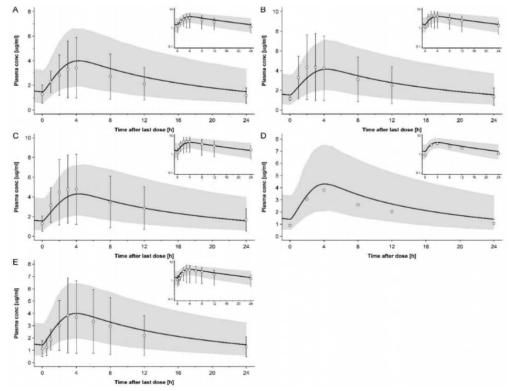


Figure 5: Plasma concentration-time profiles of dolutegravir following oral administration once a day of 50 mg in fed state in non-pregnant subjects [31,34-36].

Figure 6 illustrates the anticipated plasma concentration-time profiles of DTG during the middle and final stages of pregnancy. In Figure 7, the Goodness-of-Fit (GOF) plot displays model-predicted DTG plasma concentrations in both non-pregnant and pregnant women. Tab. 4 provides absolute values and ratios of

predicted to observed AUC 0-24, Cmax, and tmax in the pregnant populations. The model adequately captured variability, with the predicted 5th – 95th percentile range encompassing 76% of all observed concentration values in the mid-pregnancy stage and 69% in the final stage of pregnancy.

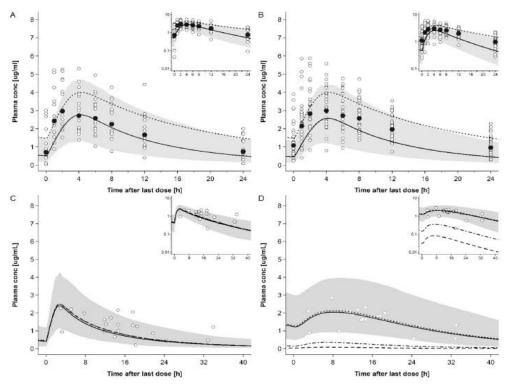


Figure 6: Plasma concentration-time profiles of dolutegravir following oral administration of 50 mg once a day in pregnant women. Semi-log scale figures are given as inset figure in the top right corners [38].

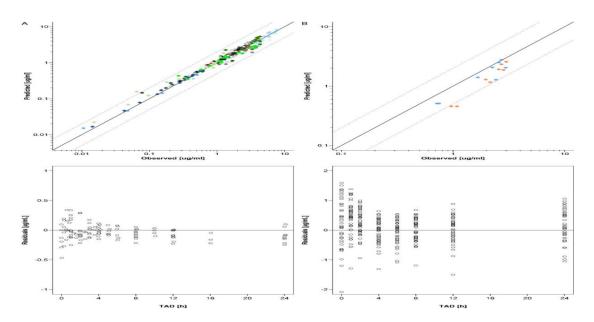


Figure 7: Goodness-of-fit (GOF) and residuals vs time plots of dolutegravir concentrations in non-pregnant subjects (A) and pregnant women (B).

A panel: Upper plot: GOF plot of geometric mean dolutegravir concentrations in non-pregnant populations. **B panel**: Upper plot: GOF plot of dolutegravir in pregnant population [28-36].²⁸⁻³⁶

6.1.2 Raltegravir

The simulated plasma concentration-time profiles of RAL in non-pregnant populations following administration of 400

mg tablet BID in fed state (i.e. the same dosing regimen as in pregnant women) are shown in Figure 8.

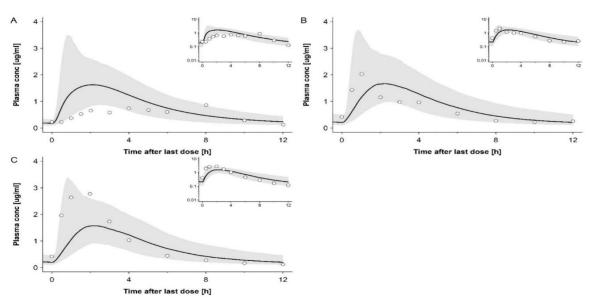


Figure 8: Plasma concentration-time profiles of raltegravir following oral administration twice a day of 400mg with moderate fat meal in non-pregnant subjects [52,53].

The predicted RAL plasma concentration-time profiles in the 2nd and 3rd trimesters of pregnancy are shown in Figure 9

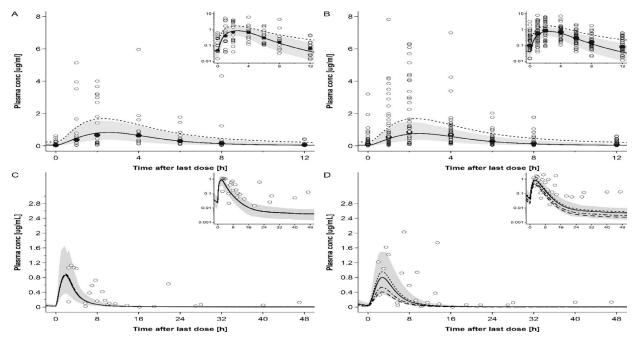


Figure 9: Plasma concentration-time profiles of raltegravir following oral administration of 400 mg twice a day in pregnant women in steady state. Semi-log scale figures are given as inset figure in the top right corners.

A: raltegravir 400 mg twice a day in pregnant women in 2nd trimester.

B: raltegravir 400 mg twice a day in pregnant women in 3rd trimester.

C: raltegravir 400 mg twice a day in pregnant women with an average gestational age of 38 weeks at delivery, represent individual concentration data in the maternal plasma.

D: raltegravir 400 mg twice a day in pregnant women with an average gestational age of 38 weeks at delivery, represent individual concentration data in the umbilical vein [39].

Figure 10 shows the GOF plot for the model-predicted RAL plasma concentrations in non-pregnant and pregnant women with the residuals versus time.

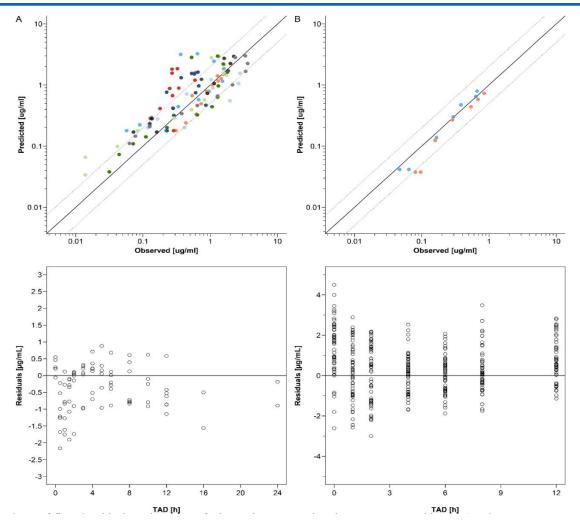


Figure 10: Goodness-of-fit and residuals vs time plots of raltegravir concentrations in non-pregnant subjects (A) and pregnant women (B).

A panel: Upper plot: GOF plot of geometric mean dolutegravir concentrations in non-pregnant populations. **B** panel: Upper plot: GOF plot of raltegravir in pregnant population [52,53].

Name of Antiviral drug and population condition	AUC _{0-t} [mg·h/L] Simulated or Predicted/Observed (Ratio)	C _{max} [mg/L] Simulate d or Predicted/Observed (Ratio)	t _{max} [h] Simulated or Predicted/Observed (Ratio)
Emtricitabine			
Nonpregnant women			
Bapuji et al ²⁰	10.60/11.20 (0.95)	3.00/2.01 (1.49)	0.90/0.78 (1.15)
Zong et al, study 1 ¹⁹	9.67/11.20 (0.86)	2.46/2.13 (1.16)	0.95/1.01 (0.94)
Zong et al, study 2 ¹⁹	9.62/11.11 (0.87)	2.36/2.17 (1.09)	0.95/1.25 (0.76)
Blum et al ¹⁸ (steady state)	10.70/10.33(1.04)	1.33/1.64 (0.81)	3.00/3.07 (0.98)
Zong et al, study 3 ¹⁹ (steady state)	9.62/9.67 (0.99)	2.44/2.11 (1.16)	1.00/1.02 (0.98)
Pregnant women GA 23-30 weeks ²¹ (steady state)	7.41/7.50 (0.99)	1.14/1.19 (0.96)	2.25/2.00 (1.13)

GA 31-35 weeks ²¹ (steady state)	7.35/6.45 (1.14)	1.17/1.32 (0.89)	2.50/2.00 (1.25)
GA 36-42 weeks ²¹ (steady state)	7.27/4.15 (1.75) ^a	1.12/0.66 (1.70) ^a	2.25/2.00 (1.13) ^a
Acyclovir			
Nonpregnant			
women			
Intravenous injection ¹²			
Study group A (2.5 mg/kg)	13.7/11.5 (1.19)		
Study group B (5.0 mg/kg)	22.8/23.3 (0.98)		
Study group C (10 mg/kg)	38.8/37.6 (1.03)		
Study group D (15 mg/kg)	56.9/44.9 (1.27)		
Oral administration ¹³			
Suspension	2.98/2.51 (1.19)	0.640/0.543 (1.18)	1.60/1.50 (1.07)
Tablet	3.09/2.50 (1.24)	0.669/0.590 (1.13)	1.65/1.75 (0.94)
Pregnant women	0.00/0.11 (0.05)	0.554/0.600.001	1 (5/0 00 (0.00)
GA 36 weeks ¹²	2.00/2.11 (0.95)	0.574/0.630 (0.91)	1.65/2.00 (0.83)
GA 38 weeks ^{12,13} (steady	2.84/3.05 (0.93)	0.726/0.791 (0.92)	1.65/1.50 (1.10)
state)			
Dolutegravir Nonpregnant women			
Castellino study ²⁸	34.0/35.9 (0.95)	2.29/2.53 (0.86)	0.95/0.50 (1.90)
Dooley study cohort 1 ²⁹	39.0/36.1 (1.08)	2.91/2.65 (1.10)	2.25/1.5 (1.50)
(steady state)	57.0/50.1 (1.00)	2.91/2.03 (1.10)	2.23/1.3 (1.30)
Dooley study cohort 2 ²⁹	41.1/42.1 (0.98)	2.96/2.91 (1.02)	2.20/2.00 (1.10)
(steady state)			
Ford study ³⁴ (steady state)	63.1/52.5(1.20)	4.00/3.43 (1.17)	4.25/4.00 (1.06)
Johnson2014, Cohort 1 (steady state) ³⁵	65.2/71.9 (0.91)	4.14/4.35 (0.95)	4.30/3.00 (1.43)
Johnson2014, Cohort2 (steady state) ³⁵	67.4/71.9 (0.94)	4.30/4.78 (0.90)	4.30/3.50 (1.23)
Song2012_high_fat ³²	66.1/83.6 (0.79)	2.97/4.19 (0.71)	4.90/5.00 (0.98)
Song2012_low_fat ³²	59.8/66.7 (0.90)	2.83/3.81 (0.74)	4.00/3.00 (1.33)
Song2012_moderate_fat ³²	64.7/71.0 (0.91)	2.94/3.86 (0.76)	4.75/4.00 (1.19)
Song2016_moderate_fat_me al ³¹ (steady state)	62.2/55.4 (1.12)	3.99/3.83 (1.04)	4.00/3.00 (1.33)
Song2013 study ³⁰ b	47.2/40.3 (1.17)	1.82/1.90 (0.96)	2.50/3.00 (0.83)
Weller study ³³	44.1/37.1 (1.19)	1.89/1.84 (1.03)	2.40/2.50 (0.96)
Wang2019 ³⁶	63.8/51.62 (1.23)	4.30/3.81(1.13)	4.10/4.00 (1.03)
Pregnant women			
2nd trimester (steady state)	34.70/42.38 (0.82)	2.77/3.00 (0.92)	4.20/2.00 (2.10)
3rd trimester (steady state)	31.91/47.59 (0.67)	2.57/3.00 (0.86)	4.20/4.00 (1.05)
Raltegravir	See OSP GitHub	See OSP GitHub	See OSP GitHub
Non-pregnant women			

Markowitz2006 ⁵⁴	8.86/7.96 (1.11)	3.04/2.24 (1.36)	0.80/1.00 (0.80)
Iwamoto2009 ⁵⁷	8.66/4.90 (1.77)	3.10/1.28 (2.42)	0.75/1.50 (0.50)
Rhee2014 ⁵⁵	9.13/8.53 (1.07)	3.11/2.22 (1.40)	0.75/2.00 (0.38)
Wenning2009 ⁵⁶	8.99/12.25 (0.73)	3.12/3.82 (0.81)	0.75/1.50 (0.50)
Brainard2011_fasted b ⁵⁸	9.66/6.47 (1.49)	3.42/1.59 (2.15)	0.75/2.00 (0.38)
Brainard2011_high fat b ⁵⁸	8.83/ 11.37 (0.78)	1.48/1.59 (0.93)	2.45/2.00 (1.23)
Brainard2011_moderate_fat b 58	8.86/6.44 (1.38)	1.54/0.74 (2.08)	2.20/4.00 (0.55)
Brainard2011_low_fat b 58	9.01/3.39 (2.66)	1.76/0.59 (2.98)	1.95/3.50 (0.56)
Taburet2015_moderate_fat_1 c ⁵⁹	8.46/8.24 (1.03)	1.65/2.03 (0.82)	2.20/1.00 (2.20)
Taburet2015_moderate_fat_2 c ⁵⁹	8.95/ 11.00 (0.81)	1.54/2.77 (0.56)	2.20/2.00 (1.10)
Pregnant women			
2nd trimester (steady state)	4.10/3.90 (1.05)	0.834/0.67 (1.22)	2.55/2.00 (1.28)
3rd trimester (steady state)	3.71/4.44 (0.84)	0.763/0.85 (0.89)	2.50/2.00 (1.25)

Table 1: provides a comparative analysis between the observed in vivo pharmacokinetic (PK) parameters and those derived from the simulated concentration-time profiles for Emtricitabine, Acyclovir, Dolutegravir, and Raltegravir, respectively.

7. Discussion

The present research study focused on the development and evaluation of maternal-fetal PBPK models for four drugs: Emtricitabine, Acyclovir, Dolutegravir, and Raltegravir. In order to assess the accuracy of these models, in vivo data was utilized, including maternal and fetal concentration data collected at different stages of pregnancy. The study found that, overall, the maternal pharmacokinetics (PK) were well-predicted by the developed models. Specifically, the observed values for Cmax, tmax, and AUC0-tlast were predicted within a 25% margin of error, as shown in Table 1. However, it should be noted that the description of interindividual variability was less satisfactory, particularly for Acyclovir. This limitation is commonly observed in current PBPK models. In the presented models, changes in drug distribution were primarily driven by increases in the volume of blood plasma and other tissues, such as fat tissue. Additionally, changes in the drug's fraction unbound also played a role. It is worth mentioning that no PK data following intravenous administration of the modeled drugs were reported, which complicates the evaluation of predicted drug distribution. Nonetheless, previous studies on pregnancy PBPK models for intravenously administered drugs, such as cefazolin, cefradine, cefuroxime, and acetaminophen, have indicated that the disposition kinetics at various stages of pregnancy were adequately predicted. This provides some confidence in the validity of the presented maternal-fetal PBPK models [24,26]. The nonpregnant model presented for intravenous administration was developed using pharmacokinetic (PK) data from men. However, it is important to note that this approach may have resulted in an underestimation of acyclovir clearance when

observed that atazanavir was concomitantly administered in 11 patients following the intake of a light meal (~360 kilocalories). This suggests that, at least in these patients, emtricitabine was taken in a fed state. As a result, the incorporation of a light meal intake was implemented in all maternal-fetal PBPK models. This adjustment led to a decrease in Cmax (maximum concentration) and an increase in tmax (time to reach maximum concentration) compared to the simulations in nonpregnant women. However, AUC0-t (area under the curve from time zero to the last measurable concentration) was not affected and remained virtually identical between predictions for the fasted and fed states in pregnant populations. These findings indicate that there may be a negative impact of food on emtricitabine PK. However, since the overall exposure is not affected, this food effect is likely of no clinical significance. It is worth noting that a small fraction of acyclovir (8.5% to 14.1%) is metabolized through the action of aldehyde dehydrogenase 2 (ALDH2). The available evidence suggests that a similar conclusion can be drawn for emtricitabine regarding its metabolism, although the specific enzyme involved in this process is not yet known. Emtricitabine undergoes two main metabolic pathways: oxidation of the thiol moiety, leading to the formation of 3r-sulfoxide diastereomers, and conjugation with glucuronic acid to form 2r-O-glucuronide. Notably, these metabolic reactions do not seem to be catalyzed by the cytochrome P450 enzyme system, as mentioned in the presented PBPK models. Interestingly, the PBPK models presented in this study indicate that the metabolic clearance of both acyclovir and emtricitabine remains unchanged during pregnancy. It is worth noting that

applied to pregnant women. In the study protocol, it was

both emtricitabine and acyclovir are substrates of renal drug transporters, implying that renal excretion plays a significant role in their elimination from the body. Emtricitabine has been found to be a substrate for efflux transporters such as MATE1, MATE2, and MRP1. However, the specific influx transporter for emtricitabine has not yet been identified. On the other hand, acyclovir is a substrate for multiple transporters, including OAT1, OAT2, OAT3, OCT1 for influx, and MATE1 and MATE2 for efflux. Unfortunately, there is currently limited evidence regarding the impact of pregnancy on the expression of these transporters. In the maternal-fetal PBPK models presented, the predicted increase in total renal clearance can be attributed to various factors, including the rise in glomerular filtration rate, kidney volume, renal blood flow, and fraction unbound. Similar to a previous study, a univariate sensitivity analysis was conducted on these parameters. Specifically, the plasma concentration-time profile for acyclovir and emtricitabine was simulated while these factors in the model were either kept constant at the nonpregnant value or adjusted to the pregnant value. The analysis conducted revealed that the observed increase in total renal clearance during pregnancy could not be solely explained by the rise in glomerular filtration rate, fraction unbound, and renal blood flow. Interestingly, it was found that the higher kidney volume played a significant role in predicting the increase in tubular net secretion, which directly influenced the tubular secretion rate and ultimately led to the observed increase in total renal clearance. Currently, there is limited information available on renal physiology and renal transporters in pregnant women, which poses challenges in identifying the specific factors contributing to the observed increase in tubular net secretion clearance. Previous pregnancy PBPK models for emtricitabine have incorporated the observed increases in net secretion clearance of metformin, a well-known substrate of OCT2, or increases in renal plasma flow to inform the rise in tubular net secretion. In the current model, a 52% increase in tubular net secretion was applied in the third trimester. However, further in vitro and in vivo data are required to uncover the underlying physiological mechanism(s) responsible for this clearance increase. In the case of acyclovir, it was observed that the predicted fetal concentrations were relatively insensitive to changes in the transfer constant but were sensitive to changes in the partition coefficient. The PBPK model used in this study slightly underestimated fetal concentrations of acyclovir due to a smaller partition coefficient, as determined from ex vivo cotyledon perfusion data, resulting in higher concentrations on the maternal side compared to the fetal side of the placenta. However, it is questionable whether the in vivo partition coefficient for acyclovir is indeed smaller than 1. It is important to note that this study focused on investigating pregnancyinduced alterations in pharmacokinetic (PK) target parameters, and further considerations on appropriate dosing regimens would require incorporating drug pharmacodynamics, which was beyond the scope of this study. It should be acknowledged that the patients included in this study, who were living with HIV, were taking multiple antiviral drugs. The clinical study used for model evaluation during pregnancy involved coadministration of emtricitabine with several other antiretroviral medications. However, it is expected that none of these co-

interactions were anticipated in this study, the therapeutic effect, such as viral load suppression, represents a combined effect of multiple antiviral drugs, making the development of a mechanistic pharmacodynamic model complex. Regarding acyclovir, the median minimum concentration (Cmin) and area under the curve (AUC) predicted throughout pregnancy consistently remained above the reported inhibitory concentration (IC50). Similarly, for emtricitabine, the median Cmin and AUC predicted throughout pregnancy were also above the reported target thresholds. However, in the lower percentiles, both Cmin and AUC during late second trimester and early third trimester were predicted to be lower than the IC50 and desired AUC, respectively. The PBPK models developed in this study were based on the population described and have not been adjusted for pharmacogenetic differences or ethnicity. However, it is not expected that there are any significant changes in this regard for emtricitabine and acyclovir. The predicted PK of DTG in pregnant, non-laboring women aligned well with clinical data obtained at different stages of pregnancy. While most PK parameters were accurately predicted, the time to reach maximum concentration (tmax) in the second trimester was slightly overestimated, and the AUCtau was underestimated in the third trimester. Further clinical studies controlling for food intake could help elucidate the reasons behind these observations. The underestimation of AUCtau in the third trimester for DTG was primarily attributed to an overestimation of total body clearance. This, combined with the results from the sensitivity analysis on UGT1A1 induction, suggests that UGT1A1 induction may be lower than initially expected, and changes in the fraction unbound are the main driver of increased total clearance, which is consistent with recent findings in the literature. The PK of RAL in pregnant, non-laboring women was generally well predicted, although variability, particularly in the first hours after drug administration, was underestimated, indicating that the model did not fully capture variability related to drug absorption. UGT1A1 plays a crucial role in the metabolism of both DTG and RAL. While in vitro experiments support an increase in UGT1A1 expression mediated by rising progesterone levels during pregnancy, there is limited information on quantitative changes in UGT1A1 expression in vivo. In this study, UGT1A1 was assumed to be induced by factors of 1.75 in the second trimester and 1.92 in the third trimester based on a previous PBPK model for acetaminophen. Maternal PK of DTG was moderately sensitive to alterations in UGT1A1 expression, while maternal PK of RAL was weakly affected. It is important to acknowledge that the limited clinical data available hindered a comprehensive evaluation of these predictions, and more data, ideally from different tissues such as maternal plasma, placenta, and umbilical vein, are needed to better assess the predictive performance of these models. However, within the limitations of this study, models that incorporated the Poulin & Theil method or the QSAR approach appeared to better predict umbilical vein concentrations compared to other models. This highlights the significance of maternal PK in fetal drug exposure and suggests that the primary elimination pathway in the fetus is transfer across the placenta back to the mother. The concentrations of RAL in maternal plasma and the umbilical vein were particularly

medications interact with emtricitabine. Although no drug-drug

underestimated after 12 hours, which corresponds to the dosing interval. One of the reasons for the underestimation of maternal plasma concentrations of RAL at delivery seems to be the fast absorption assumed in the model. While the PBPK models for non-laboring women considered a fed state to reflect the lack of food restrictions in the clinical study, the PBPK model for laboring women assumed a fasted state (as it is unlikely for women to consume food right before labor and delivery). However, it was noted that the PBPK model at delivery still accounted for delayed gastric passage of RAL (and DTG) due to evidence suggesting that gastric emptying and drug absorption from the gastrointestinal tract are slowed during labor. On May 18th, 2018, the US FDA issued a warning letter regarding DTG, stating that it may cause serious birth defects involving the brain, spine, and spinal cord. These preliminary findings were observed in a study conducted in Botswana, where women who received DTG at the time of conception were affected. The exact mechanism behind the teratogenicity of DTG is not yet fully understood, but one hypothesis suggests that it interferes with folic acid binding to the folate receptor α , leading to reduced levels of folic acid in the fetus. Since folic acid is crucial for neural tube development, a decrease in folic acid levels could potentially result in neural tube defects in the fetus. In vitro results presented by Zamek et al. indicate that free DTG concentrations of approximately 37 µM correspond to a 36% inhibition of the folate receptor α. To provide an in vivo context, the PBPK model was extrapolated to the 6th gestational week, assuming a 33% induction of UGT1A1 in the first trimester. Unbound DTG concentrations were then predicted in the maternal blood of the placenta, with a maximum predicted concentration of 0.06 µM at steady state during the 6th gestational week. Using an Emax model fitted to the data reported by Zamek et al., where Emax was 1.0 and EC50 was 1276 µM, this DTG concentration corresponds to an inhibition of the folate receptor α by approximately 7%. However, it is important to note that these predicted concentrations cannot be evaluated due to the lack of clinical data, and therefore should not be used to guide dosing decisions. This example highlights the potential theoretical contribution of PBPK modeling in supporting decision-making regarding the use of DTG during pregnancy. However, it also emphasizes the need for clinical data to validate and enhance the reliability of model-based predictions.

8. Conclusions

In summary, the developed maternal-fetal PBPK models have successfully predicted the pharmacokinetic profiles of Emtricitabine, Acyclovir, Dolutegravir, and Raltegravir at various stages of pregnancy. This enhances confidence in utilizing one of the key strengths of PBPK analyses, which is the ability to extrapolate drug pharmacokinetics from well-characterized populations of healthy adults to the unique population of pregnant women. The investigation into pregnancy-induced changes in pharmacokinetic target parameters confirms the appropriateness of current dosing regimens for acyclovir and emtricitabine, at least for the average pregnant patient. The presented model strengthens the confidence in such models, which is crucial when applying them to inform the design of clinical trials for drugs with similar pharmacokinetic characteristics in pregnant

women. Given the limited participation of pregnant women in clinical trials, PBPK modeling can serve as a valuable tool to supplement the understanding of pharmacokinetics in cases where clinical data is sparse or unavailable. While these models should not be seen as a substitute for clinical trials, they contribute to a broader and mechanistic understanding of pharmacokinetics, with the potential to enhance drug safety and efficacy for both the mother and the fetus [54-83].

References

- Haas, D. M., Marsh, D. J., Dang, D. T., Parker, C. B., Wing, D. A., Simhan, H. N., ... & Reddy, U. M. (2018). Prescription and other medication use in pregnancy. *Obstetrics & Gynecology*, 131(5), 789-798.
- Straface, G., Selmin, A., Zanardo, V., De Santis, M., Ercoli, A., & Scambia, G. (2012). Herpes simplex virus infection in pregnancy. *Infectious Diseases in Obstetrics and Gynecology*, 2012(1), 385697.
- Ending, A. I. D. S. (2019). Progress towards 90-90-90 targets. Joint United Nations Programme on HIV/AIDS (UNAIDS).
- Dallmann, A., Pfister, M., van den Anker, J., & Eissing, T. (2018). Physiologically based pharmacokinetic modeling in pregnancy: a systematic review of published models. *Clinical Pharmacology & Therapeutics*, 104(6), 1110-1124.
- Zhang, Z., Imperial, M. Z., Patilea-Vrana, G. I., Wedagedera, J., Gaohua, L., & Unadkat, J. D. (2017). Development of a novel maternal-fetal physiologically based pharmacokinetic model I: insights into factors that determine fetal drug exposure through simulations and sensitivity analyses. *Drug Metabolism and Disposition*, 45(8), 920-938.
- 6. Dallmann, A., Solodenko, J., Ince, I., & Eissing, T. (2018). Applied concepts in PBPK modeling: how to extend an open systems pharmacology model to the special population of pregnant women. *CPT: pharmacometrics & systems pharmacology*, 7(7), 419-431.
- Liu, X. I., Momper, J. D., Rakhmanina, N., van den Anker, J. N., Green, D. J., Burckart, G. J., ... & Dallmann, A. (2020). Physiologically based pharmacokinetic models to predict maternal pharmacokinetics and fetal exposure to emtricitabine and acyclovir. *The Journal of Clinical Pharmacology*, 60(2), 240-255.
- de Miranda, P., Good, S. S., Laskin, O. L., Krasny, H. C., Connor, J. D., & Lietman, P. S. (1981). Disposition of intravenous radioactive acyclovir. *Clinical Pharmacology* & *Therapeutics*, 30(5), 662-672.
- Cheng, Y., Vapurcuyan, A., Shahidullah, M., Aleksunes, L. M., & Pelis, R. M. (2012). Expression of organic anion transporter 2 in the human kidney and its potential role in the tubular secretion of guanine-containing antiviral drugs. *Drug Metabolism and Disposition*, 40(3), 617-624.
- De Miranda, P., & Good, S. S. (1992). Species differences in the metabolism and disposition of antiviral nucleoside analogues: 1. Acyclovir. *Antiviral Chemistry and Chemotherapy*, 3(1), 1-8.
- 11. Gunness, P., Aleksa, K., Bend, J., & Koren, G. (2011). Acyclovir-induced nephrotoxicity: the role of the acyclovir aldehyde metabolite. *Translational Research*, 158(5), 290-

301.

- Laskin, O. L., Longstreth, J. A., Saral, R. E. I. N., De Miranda, P., Keeney, R., & Lietman, P. S. (1982). Pharmacokinetics and tolerance of acyclovir, a new anti-herpesvirus agent, in humans. *Antimicrobial agents and chemotherapy*, 21(3), 393-398.
- Palma-Aguirre, J. A., Absalón-Reyes, J. A., Novoa-Heckel, G., de Lago, A., Oliva, I., Rodríguez, Z., ... & Namur, S. (2007). Bioavailability of two oral suspension and two oral tablet formulations of acyclovir 400 mg: two single-dose, open-label, randomized, two-period crossover comparisons in healthy Mexican adult subjects. *Clinical therapeutics*, 29(6), 1146-1152.
- Frenkel, L. M., Brown, Z. A., Bryson, Y. J., Corey, L., Unadkat, J. D., Hensleigh, P. A., ... & Connor, J. D. (1991). Pharmacokinetics of acyclovir in the term human pregnancy and neonate. *American journal of obstetrics and* gynecology, 164(2), 569-576.
- Kimberlin, D. F., Weller, S., Whitley, R. J., Andrews, W. W., Hauth, J. C., Lakeman, F., & Miller, G. (1998). Pharmacokinetics of oral valacyclovir and acyclovir in late pregnancy. *American journal of obstetrics and gynecology*, 179(4), 846-851.
- Leung, D. T., Henning, P. A., Wagner, E. C., Blasig, A., Wald, A., Sacks, S. L., ... & Money, D. M. (2009). Inadequacy of plasma acyclovir levels at delivery in patients with genital herpes receiving oral acyclovir suppressive therapy in late pregnancy. *Journal of obstetrics and Gynaecology Canada*, *31*(12), 1137-1143.
- 17. Bartlett, J. (2004). Recommendations for use of antiretroviral drugs in pregnant hiv-1-infected women for maternal health and interventions to reduce perinatal hiv-1 transmission in the United States. *Infectious Diseases in Clinical Practice*, *12*(6), 363-364.
- Blum, M. R., Chittick, G. E., Begley, J. A., & Zong, J. (2007). Steady-state pharmacokinetics of emtricitabine and tenofovir disoproxil fumarate administered alone and in combination in healthy volunteers. *The Journal of Clinical Pharmacology*, 47(6), 751-759.
- Zong, J., Chittick, G. E., Wang, L. H., Hui, J., Begley, J. A., & Blum, M. R. (2007). Pharmacokinetic evaluation of emtricitabine in combination with other nucleoside antivirals in healthy volunteers. *The Journal of Clinical Pharmacology*, 47(7), 877-889.
- Bapuji, A. T., Nagesh, M., Ramaraju, D., Syedba, S., Kiran, R., & Ravinder, S. (2010). A bioequivalence study comparing two formulation of emtricitabine capsules. J Bioequiv Availab, 2, 11-14.
- Stek, A. M., Best, B. M., Luo, W., Capparelli, E., Burchett, S., Hu, C., ... & Mirochnick, M. (2012). Effect of pregnancy on emtricitabine pharmacokinetics. *HIV medicine*, 13(4), 226-235.
- De Sousa Mendes, M., Hirt, D., Vinot, C., Valade, E., Lui, G., Pressiat, C., ... & Benaboud, S. (2016). Prediction of human fetal pharmacokinetics using ex vivo human placenta perfusion studies and physiologically based models. *British journal of clinical pharmacology*, *81*(4), 646-657.
- 23. Reese, M. J., Savina, P. M., Generaux, G. T., Tracey, H.,

Humphreys, J. E., Kanaoka, E., ... & Polli, J. W. (2013). In vitro investigations into the roles of drug transporters and metabolizing enzymes in the disposition and drug interactions of dolutegravir, a HIV integrase inhibitor. *Drug Metabolism and Disposition*, *41*(2), 353-361.

- 24. Dallmann, A., Ince, I., Meyer, M., Willmann, S., Eissing, T., & Hempel, G. (2017). Gestation-specific changes in the anatomy and physiology of healthy pregnant women: an extended repository of model parameters for physiologically based pharmacokinetic modeling in pregnancy. *Clinical pharmacokinetics*, 56, 1303-1330.
- Dallmann, A., Ince, I., Coboeken, K., Eissing, T., & Hempel, G. (2018). A physiologically based pharmacokinetic model for pregnant women to predict the pharmacokinetics of drugs metabolized via several enzymatic pathways. *Clinical pharmacokinetics*, 57, 749-768.
- 26. Mian, P., Van Den Anker, J. N., van Calsteren, K., Annaert, P., Tibboel, D., Pfister, M., ... & Dallmann, A. (2020). Physiologically based pharmacokinetic modeling to characterize acetaminophen pharmacokinetics and N-acetyl-p-benzoquinone imine (NAPQI) formation in nonpregnant and pregnant women. *Clinical pharmacokinetics*, 59, 97-110.
- 27. Center For Drug Evaluation and Research: Clinical Pharmacology and Biopharmaceutics Review (s). Access at: (accessed on October 8, 2019).
- Castellino, S., Moss, L., Wagner, D., Borland, J., Song, I., Chen, S., ... & Savina, P. M. (2013). Metabolism, excretion, and mass balance of the HIV-1 integrase inhibitor dolutegravir in humans. *Antimicrobial agents and chemotherapy*, 57(8), 3536-3546.
- 29. Dooley, K. E., Sayre, P., Borland, J., Purdy, E., Chen, S., Song, I., ... & Flexner, C. (2013). Safety, tolerability, and pharmacokinetics of the HIV integrase inhibitor dolutegravir given twice daily with rifampin or once daily with rifabutin: results of a phase 1 study among healthy subjects. *JAIDS Journal of Acquired Immune Deficiency Syndromes*, 62(1), 21-27.
- Song, I. H., Borland, J., Savina, P. M., Chen, S., Patel, P., Wajima, T., ... & Piscitelli, S. C. (2013). Pharmacokinetics of single-dose dolutegravir in HIV-seronegative subjects with moderate hepatic impairment compared to healthy matched controls. *Clinical pharmacology in drug development*, 2(4), 342-348.
- Song, I., Weller, S., Patel, J., Borland, J., Wynne, B., Choukour, M., ... & Piscitelli, S. (2016). Effect of carbamazepine on dolutegravir pharmacokinetics and dosing recommendation. *European journal of clinical pharmacology*, 72, 665-670.
- 32. Song, I., Borland, J., Chen, S., Patel, P., Wajima, T., Peppercorn, A., & Piscitelli, S. C. (2012). Effect of food on the pharmacokinetics of the integrase inhibitor dolutegravir. *Antimicrobial agents and chemotherapy*, *56*(3), 1627-1629.
- Weller, S., Borland, J., Chen, S., Johnson, M., Savina, P., Wynne, B., ... & Piscitelli, S. C. (2014). Pharmacokinetics of dolutegravir in HIV-seronegative subjects with severe renal impairment. *European journal of clinical pharmacology*, 70, 29-35.

- 34. Ford, S. L., Gould, E., Chen, S., Margolis, D., Spreen, W., Crauwels, H., & Piscitelli, S. (2013). Lack of pharmacokinetic interaction between rilpivirine and integrase inhibitors dolutegravir and GSK1265744. *Antimicrobial agents and chemotherapy*, 57(11), 5472-5477.
- Johnson, M., Borland, J., Chen, S., Savina, P., Wynne, B., & Piscitelli, S. (2014). Effects of boceprevir and telaprevir on the pharmacokinetics of dolutegravir. *British journal of clinical pharmacology*, 78(5), 1043-1049.
- Wang, X., Cerrone, M., Ferretti, F., Castrillo, N., Maartens, G., McClure, M., & Boffito, M. (2019). Pharmacokinetics of dolutegravir 100 mg once daily with rifampicin. *International journal of antimicrobial agents*, 54(2), 202-206.
- 37. Min, S., Song, I., Borland, J., Chen, S., Lou, Y., Fujiwara, T., & Piscitelli, S. C. (2010). Pharmacokinetics and safety of S/GSK1349572, a next-generation HIV integrase inhibitor, in healthy volunteers. *Antimicrobial agents and chemotherapy*, 54(1), 254-258.
- Mulligan, N., Best, B. M., Wang, J., Capparelli, E. V., Stek, A., Barr, E., ... & Mirochnick, M. (2018). Dolutegravir pharmacokinetics in pregnant and postpartum women living with HIV. *Aids*, *32*(6), 729-737.
- Watts, D. H., Stek, A., Best, B. M., Wang, J., Capparelli, E. V., Cressey, T. R., ... & IMPAACT 1026s Study Team. (2014). Raltegravir pharmacokinetics during pregnancy. *JAIDS Journal of Acquired Immune Deficiency Syndromes*, 67(4), 375-381.
- Waitt, C., Orrell, C., Walimbwa, S., Singh, Y., Kintu, K., Simmons, B., ... & Khoo, S. (2019). Safety and pharmacokinetics of dolutegravir in pregnant mothers with HIV infection and their neonates: a randomised trial (DolPHIN-1 study). *PLoS medicine*, 16(9), e1002895.
- Blonk, M. I., Colbers, A. P., Hidalgo-Tenorio, C., Kabeya, K., Weizsäcker, K., Haberl, A. E., ... & Wood, C. (2015). Raltegravir in HIV-1–Infected pregnant women: pharmacokinetics, safety, and efficacy. *Clinical infectious diseases*, *61*(5), 809-816.
- 42. Dallmann, A., Ince, I., Solodenko, J., Meyer, M., Willmann, S., Eissing, T., & Hempel, G. (2017). Physiologically based pharmacokinetic modeling of renally cleared drugs in pregnant women. *Clinical pharmacokinetics*, *56*, 1525-1541.
- Zhang, Z., & Unadkat, J. D. (2017). Development of a novel maternal-fetal physiologically based pharmacokinetic model II: verification of the model for passive placental permeability drugs. *Drug Metabolism and Disposition*, 45(8), 939-946.
- Grießinger, J. A., Hauptstein, S., Laffleur, F., Netsomboon, K., & Bernkop-Schnürch, A. (2016). Evaluation of the impact of multivalent metal ions on the permeation behavior of Dolutegravir sodium. *Drug development and industrial pharmacy*, 42(7), 1118-1126.
- 45. Moss, D. M., Kwan, W. S., Liptrott, N. J., Smith, D. L., Siccardi, M., Khoo, S. H., ... & Owen, A. (2011). Raltegravir is a substrate for SLC22A6: a putative mechanism for the interaction between raltegravir and tenofovir. *Antimicrobial agents and chemotherapy*, *55*(2), 879-887.

- 46. Open Systems Pharmacology: Compounds: Definition and Work Flows.
- Poulin, P., Schoenlein, K., & Theil, F. P. (2001). Prediction of adipose tissue: plasma partition coefficients for structurally unrelated drugs. *Journal of pharmaceutical sciences*, 90(4), 436-447.
- Poulin, P., & Theil, F. P. (2002). Prediction of pharmacokinetics prior to in vivo studies. 1. Mechanismbased prediction of volume of distribution. *Journal of pharmaceutical sciences*, 91(1), 129-156.
- 49. Rodgers, T., Leahy, D., & Rowland, M. (2005). Physiologically based pharmacokinetic modeling 1: predicting the tissue distribution of moderate-to-strong bases. *Journal of pharmaceutical sciences*, 94(6), 1259-1276.
- Rodgers, T., & Rowland, M. (2006). Physiologically based pharmacokinetic modelling 2: predicting the tissue distribution of acids, very weak bases, neutrals and zwitterions. *Journal of pharmaceutical sciences*, 95(6), 1238-1257.
- Kassahun, K., McIntosh, I., Cui, D., Hreniuk, D., Merschman, S., Lasseter, K., ... & Wenning, L. A. (2007). Metabolism and disposition in humans of raltegravir (MK-0518), an anti-AIDS drug targeting the human immunodeficiency virus 1 integrase enzyme. *Drug Metabolism and Disposition*, 35(9), 1657-1663.
- 52. Markowitz, M., Morales-Ramirez, J. O., Nguyen, B. Y., Kovacs, C. M., Steigbigel, R. T., Cooper, D. A., ... & Protocol 004 Study Team. (2006). Antiretroviral activity, pharmacokinetics, and tolerability of MK-0518, a novel inhibitor of HIV-1 integrase, dosed as monotherapy for 10 days in treatment-naive HIV-1-infected individuals. *JAIDS Journal of Acquired Immune Deficiency Syndromes*, 43(5), 509-515.
- 53. Rhee, E. G., Rizk, M. L., Brainard, D. M., Gendrano III, I. N., Jin, B., Wenning, L. A., ... & Iwamoto, M. (2014). A pharmacokinetic comparison of adult and paediatric formulations of raltegravir in healthy adults.
- Laufer, R., Paz, O. G., Di Marco, A., Bonelli, F., Monteagudo, E., Summa, V., & Rowley, M. (2009). Quantitative prediction of human clearance guiding the development of Raltegravir (MK-0518, isentress) and related HIV integrase inhibitors. *Drug metabolism and disposition*, 37(4), 873-883.
- Takaku, T., Nagahori, H., Sogame, Y., & Takagi, T. (2015). Quantitative structure–activity relationship model for the fetal–maternal blood concentration ratio of chemicals in humans. *Biological and Pharmaceutical Bulletin, 38*(6), 930-934.
- Wenning, L. A., Hanley, W. D., Brainard, D. M., Petry, A. S., Ghosh, K., Jin, B., ... & Iwamoto, M. (2009). Effect of rifampin, a potent inducer of drug-metabolizing enzymes, on the pharmacokinetics of raltegravir. *Antimicrobial agents and chemotherapy*, 53(7), 2852-2856.
- Iwamoto, M., Wenning, L. A., Nguyen, B. Y., Teppler, H., Moreau, A. R., Rhodes, R. R., ... & Wagner, J. A. (2009). Effects of omeprazole on plasma levels of raltegravir. *Clinical infectious diseases*, 48(4), 489-492.
- 58. Brainard, D. M., Friedman, E. J., Jin, B., Breidinger, S. A.,

Tillan, M. D., Wenning, L. A., ... & Iwamoto, M. (2010). Effect of low-, moderate-, and high-fat meals on raltegravir pharmacokinetics. *Journal of clinical pharmacology*, *51*(3), 422-427.

- Taburet, A. M., Sauvageon, H., Grinsztejn, B., Assuied, A., Veloso, V., Pilotto, J. H., ... & Molina, J. M. (2015). Pharmacokinetics of raltegravir in HIV-infected patients on rifampicin-based antitubercular therapy. *Clinical Infectious Diseases, 61*(8), 1328-1335.
- Dallmann, A., Mian, P., den Anker, J. V., & Allegaert, K. (2019). Clinical pharmacokinetic studies in pregnant women and the relevance of pharmacometric tools. *Current pharmaceutical design*, 25(5), 483-495.
- Ito, S., Kusuhara, H., Kuroiwa, Y., Wu, C., Moriyama, Y., Inoue, K., ... & Sugiyama, Y. (2010). Potent and specific inhibition of mMate1-mediated efflux of type I organic cations in the liver and kidney by pyrimethamine. *Journal* of Pharmacology and Experimental Therapeutics, 333(1), 341-350.
- Tanihara, Y., Masuda, S., Sato, T., Katsura, T., Ogawa, O., & Inui, K. I. (2007). Substrate specificity of MATE1 and MATE2-K, human multidrug and toxin extrusions/H+organic cation antiporters. *Biochemical pharmacology*, 74(2), 359-371.
- 63. Bousquet, L., Pruvost, A., Didier, N., Farinotti, R., & Mabondzo, A. (2008). Emtricitabine: inhibitor and substrate of multidrug resistance associated protein. *European journal of pharmaceutical sciences*, *35*(4), 247-256.
- 64. Takeda, M., Khamdang, S., Narikawa, S., Kimura, H., Kobayashi, Y., Yamamoto, T., ... & Endou, H. (2002). Human organic anion transporters and human organic cation transporters mediate renal antiviral transport. *Journal* of Pharmacology and Experimental Therapeutics, 300(3), 918-924.
- Ye, J., Liu, Q., Wang, C., Meng, Q., Peng, J., Sun, H., ... & Liu, K. (2012). Inhibitory effect of JBP485 on renal excretion of acyclovir by the inhibition of OAT1 and OAT3. *European journal of pharmaceutical sciences*, 47(2), 341-346.
- 66. Cheung, K. L., & Lafayette, R. A. (2013). Renal physiology of pregnancy. *Advances in chronic kidney disease*, 20(3), 209-214.
- Race, J. E., Grassl, S. M., Williams, W. J., & Holtzman, E. J. (1999). Molecular cloning and characterization of two novel human renal organic anion transporters (hOAT1 and hOAT3). *Biochemical and biophysical research communications*, 255(2), 508-514.
- Lee, N., Hebert, M. F., Prasad, B., Easterling, T. R., Kelly, E. J., Unadkat, J. D., & Wang, J. (2013). Effect of gestational age on mRNA and protein expression of polyspecific organic cation transporters during pregnancy. *Drug Metabolism and Disposition*, 41(12), 2225-2232.
- Pascolo, L., Fernetti, C., Pirulli, D., Crovella, S., Amoroso, A., & Tiribelli, C. (2003). Effects of maturation on RNA transcription and protein expression of four MRP genes in human placenta and in BeWo cells. *Biochemical and biophysical research communications*, 303(1), 259-265.
- 70. Bouazza, N., Foissac, F., Hirt, D., Urien, S., Benaboud,

S., Lui, G., & Treluyer, J. M. (2019). Methodological approaches to evaluate fetal drug exposure. *Current pharmaceutical design*, 25(5), 496-504.

- Department of Health and Human Services: Guidelines for the use of antiretroviral agents in adults and adolescents living with HIV; drug-drug interactions. Accessed September 12, 2018.
- 72. Khatri, A., Dutta, S., Dunbar, M., Podsadecki, T., Trinh, R., Awni, W., & Menon, R. (2016). Evaluation of drugdrug interactions between direct-acting anti-hepatitis C virus combination regimens and the HIV-1 antiretroviral agents raltegravir, tenofovir, emtricitabine, efavirenz, and rilpivirine. *Antimicrobial agents and chemotherapy*, 60(5), 2965-2971.
- Amini, H., Casamassimo, P. S., Lin, H. L., & Hayes, J. R. (2008). Advocacy training in US advanced pediatric dentistry training programs. *Pediatric dentistry*, 30(2), 141-146.
- 74. Xia, B., Heimbach, T., Gollen, R., Nanavati, C., & He, H. (2013). A simplified PBPK modeling approach for prediction of pharmacokinetics of four primarily renally excreted and CYP3A metabolized compounds during pregnancy. *The AAPS journal*, 15, 1012-1024.
- 75. De Sousa Mendes, M., Hirt, D., Urien, S., Valade, E., Bouazza, N., Foissac, F., ... & Benaboud, S. (2015). Physiologically-based pharmacokinetic modeling of renally excreted antiretroviral drugs in pregnant women. *British journal of clinical pharmacology*, 80(5), 1031-1041.
- 76. Bollen, P., Freriksen, J., Konopnicki, D., Weizsäcker, K., Hidalgo Tenorio, C., Moltó, J., ... & Pharmacokinetics of ANtiretroviral agents in HIV-infected pregNAnt women Network Nellen J Lyons F Lambert J Wyen C Faetkenheuer G Rockstroh JK Schwarze-Zander C Tariq Sadiq S Gilleece Y Wood C Schalkwijk S Bukkems V. (2021). The effect of pregnancy on the pharmacokinetics of total and unbound dolutegravir and its main metabolite in women living with human immunodeficiency virus. *Clinical Infectious Diseases, 72*(1), 121-127.
- Jeong, H. Y. U. N. Y. O. U. N. G., Choi, S. U. Y. O. U. N. G., Song, J. W., Chen, H. U. I. Q. I. N. G., & Fischer, J. H. (2008). Regulation of UDP-glucuronosyltransferase (UGT) 1A1 by progesterone and its impact on labetalol elimination. *Xenobiotica*, 38(1), 62-75.
- Bataille, A., Rousset, J., Marret, E., & Bonnet, F. (2014). Ultrasonographic evaluation of gastric content during labour under epidural analgesia: a prospective cohort study. *British Journal of Anaesthesia*, 112(4), 703-707.
- Davison, J. S., Davison, M. C., & Hay, D. M. (1970). Gastric emptying time in late pregnancy and labour. J Obstet Gynaecol Br Commonw, 37-41.
- Whitehead, E. M., Smith, M., Dean, Y., & O'sullivan, G. (1993). An evaluation of gastric emptying times in pregnancy and the puerperium. *Anaesthesia*, 48(1), 53-57.
- FDA, U. (2018). FDA drug safety communication: FDA to evaluate potential risk of neural tube birth defects with HIV medicine dolutegravir (Juluca, Tivicay, Triumeq); 2018.
- 82. WHO: Potential safety issue affecting women living with HIV using dolutegravir at the time of conception. Accessat

: Statement_on_DTG_18May_2018final.pdf (accessed on November 11, 2019). 2018;

83. Zamek-Gliszczynski, M. J., Zhang, X., Mudunuru, J., Du, Y., Chen, J. L., Taskar, K. S., ... & Romach, E. H. (2019). Clinical extrapolation of the effects of dolutegravir and other HIV integrase inhibitors on folate transport pathways. *Drug Metabolism and Disposition*, *47*(8), 890-898.

Copyright: ©2024 Mohamed Abdulsamed, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.